Prevalence of Vibrio Cholerae in Northern Nigeria

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Abstract

One hundred and fifty six (156) stool samples were analyzed using selective cultural methods for the isolation of Vibrio species. They were from epidemic areas namely; Jos (Plateau State), Argungu (Kebbi State), Kwali and Gwagwa (Abuja) and Suleja (Niger State) all in Northern Nigeria. 123 (78.8%) were males and 33 (21.1%) females.

Eight of these samples were positive for Vibrio cholera with the distribution as follows; Jos – 2, Argungu – 5 and Kwali – 1.

Serotyping showed that all were of the Ogawa serotype.

Antibiotic sensitivity pattern showed that all eight (100%) samples were sensitive to Ceftazidime, Cefuroxime Axetil, Gentamycin, Sparfloxacin and Tetracycline, 5 (62.5%) sensitive to Erythromycin and 4 (50%) sensitive to Chloramphenicol. The antibiotics that all the isolates were resistant to were Ampicillin, Cloxacillin, Streptomycin and Penicillin.

The study showed that Vibrio cholerae is one of the causes of gastroenteritis epidemics in Northern Nigeria.

KEY WORDS: PREVALENCE, VIBRIO CHOLEREA, NORTHERN NIGERIA

Introduction

Vibrio cholerae is usually the cause of several gastrointestinal epidemics that breakout in the Northern Nigeria. This can be attributed to several factors which include; standard of living, poor sewage management, poor state of personal hygiene, unavailability portable water and occasional floods.

The particular strain, serogroup, and antibiogram of the Vibrio cholerae that cause epidemics in these areas are rarely available. The aims and objectives of this research are:

(i) To isolate the Vibrio species responsible for cholera outbreaks in the Northern Nigeria.

(ii) To determine the most prevalent of the serotypes

(iii) To determine the antibiogram of the isolates.

Methodology

The study area was Northern region of Nigeria, West Africa. The region consists of nineteen(19) states together with The Federal Capital Territory Abuja. Occasional outbreaks have been reported in this area year after year. The vegetation in the region is Guinea savannah with moderate rainfall recorded (WHO 2009).

Purposive sampling method was used arriving at a sample size of 156 stool samples which were collected from in and out patients of both sexes and different age groups in five different locations with reported cases of gastro enteritis in 2010/2011 namely; Argungu (Kebbi State), Jos (Plateau State), Suleja (Niger State), Kwali and Gwagwa (FCT).

Stool samples in epidemic areas of gastroenteritis were collected in sterile, clean, wide mouthed, screw cap bottles, patients on chemotherapy treatment and yet to be treated ones were considered.

Samples were processed according to the guidelines on laboratory method for the diagnosis of Vibrio cholera by the Centre for Disease Control (CDC) which includes; Macros copy, Microscopy, Culture, Biochemical testing, Serology and Antimicrobial susceptibility testing.

For culture, samples were inoculated into 10mls of Alkaline Peptone Water (APW) and also directly onto Thiosulphate Citrate Bile salt Sucrose Agar (TCBS) and incubated for 24
hours at 37°C. After 6 hours of incubation, subcultures were made from the surface growth on the APW onto TCBS and incubated overnight at 37°C.

After overnight incubation, suspected colonies appear as large as 2-4 mm in diameter, slightly flattened, yellow as a result of sucrose fermentation with opaque centre and translucent periphery. Gram staining was done on them and read under ×100 objective, Gram negative straight or comma shaped colonies were further identified.

Suspicious colonies from TCBS Agar were picked and inoculated into Alkaline Peptone Water, Nutrient Agar (N.A.) and Mac Conkey Agar. Identities of the isolates were determined by biochemical and serological tests, the serological tests performed on the samples were:

Oxidase Test, Biochemical Tests like, Nitroso Indole Test, Citrate Utilization Test, Sugar Fermentation Test, Urease Test and Serotyping

Antibiotic susceptibility testing was done using Agar disc diffusion method. The following discs were used; Ampicillin 12µg, Chloramphenicol 10µg, Tetracycline 10µg, Streptomycin 10µg, Cotrimoxazole 25µg, Erythromycin 5µg, Ofloxacin 5µg, Ciprofloxacin 30µg, Ceftaxine 30µg and Gentamicin 10µg.

Zone of inhibition ≥25mm was confirmed sensitive, ≥18mm - moderately sensitive, 13-17mm - intermediate and ≤12mm - resistant.

Results

Stool samples were collected from one hundred and fifty six (156) people from epidemic areas (Jos, Argungu, Kwali, Gwagwa and Suleja). The number of samples collected from each location were as follows; Jos -36(23%), Argungu – 27(17.3%), Kwali – 43(27.6%), Gwagwa – 35(22.4%) and Suleja -15(9.6%) samples each. This is made up of 123 (78.8%) male and 33 (21.1%) female patients (Table 1).

Eight (5.13%) of these samples were positive for *Vibrio cholerae* and the distribution was as follows; Jos – 2(25%), Argungu – 5(62.5%) and Kwali – 1(12.5%) [Table 2].

The distribution of the positive samples with regards to in and out patients were 5(62.5%) in-patients and 3(37.5%) out-patients.

All (100%) of the *Vibrio Species* isolated were Ogawa serotype (Table 3).

The antibiotic sensitivity pattern of the isolates to various antibiotics is as follows; all the 8 (100%) were sensitive to Cefuroxime Axetil, Ceftazidime, Gentamycin, Sparfloxacine and Tetracycline, Five (62.5%) were sensitive to Erythromycin and 4 (50%) to Chloramphenicol. All were resistant to Ampicillin, Cloxacillin, Streptomycin and Penicillin.

The relationship between isolation rate and the length of stay of the patients in the hospital showed that 7 (87.5%) of the patients stayed less than 2 days in the hospital, while only 1 (12.5%) had stayed up to 2 days in the hospital.

**Table 1:** Sources of sample/ Number of samples collected/sex of patient

<table>
<thead>
<tr>
<th>Source of sample</th>
<th>Number of samples</th>
<th>Sex</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Jos</td>
<td>36</td>
<td>Male</td>
<td>30</td>
</tr>
<tr>
<td>Argungu</td>
<td>27</td>
<td>Male</td>
<td>16</td>
</tr>
<tr>
<td>Kwali</td>
<td>43</td>
<td>Male</td>
<td>40</td>
</tr>
<tr>
<td>Gwagwa</td>
<td>35</td>
<td>Male</td>
<td>24</td>
</tr>
<tr>
<td>Suleja</td>
<td>15</td>
<td>Male</td>
<td>13</td>
</tr>
<tr>
<td>Total</td>
<td>156</td>
<td>Male</td>
<td>123</td>
</tr>
</tbody>
</table>

**Table 2:** Sources of Sample, Number Collected in Relation to Sex and Number/ Percentage of Isolates.

<table>
<thead>
<tr>
<th>Source of sample</th>
<th>No. of samples</th>
<th>No. of isolates</th>
<th>% Yielding</th>
</tr>
</thead>
<tbody>
<tr>
<td>Jos</td>
<td>36</td>
<td>2(5.4)</td>
<td></td>
</tr>
<tr>
<td>Argungu</td>
<td>27</td>
<td>5(18.5)</td>
<td></td>
</tr>
</tbody>
</table>
Table 3: Age Group of Patients in Relation to the Sources of Samples and Number of Isolates.

<table>
<thead>
<tr>
<th>Age group</th>
<th>Jos</th>
<th>Argungu</th>
<th>Kwali</th>
<th>Gwagwa</th>
<th>Suleja</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>≤9</td>
<td>8</td>
<td>3(1)</td>
<td>1</td>
<td>2</td>
<td>1</td>
<td>36</td>
</tr>
<tr>
<td>10-19</td>
<td>8(1)</td>
<td>9(1)</td>
<td>3</td>
<td>3</td>
<td>4</td>
<td>27</td>
</tr>
<tr>
<td>20-29</td>
<td>4(1)</td>
<td>12(3)</td>
<td>27(1)</td>
<td>9</td>
<td>2</td>
<td>43</td>
</tr>
<tr>
<td>30-39</td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>16</td>
<td>2</td>
<td>35</td>
</tr>
<tr>
<td>40-49</td>
<td>10</td>
<td>0</td>
<td>9</td>
<td>5</td>
<td>3</td>
<td>15</td>
</tr>
<tr>
<td>50-59</td>
<td>3</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>3</td>
</tr>
<tr>
<td>60-69</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>≥70</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Total</td>
<td>36</td>
<td>27</td>
<td>43</td>
<td>35</td>
<td>15</td>
<td>156</td>
</tr>
</tbody>
</table>

Discussion

In this study, one hundred and fifty six (156) stool samples were analyzed using selective cultural methods for the isolation of *Vibrio cholerae*. Only 8 (5.13%) of the 156 samples yielded *Vibrio cholerae*. This shows that *Vibrio cholerae* is associated with and a cause of some gastroenteritis outbreaks but not the sole causative agent of these outbreaks in Northern Nigeria in agreement with the work of Umoh, et al (2003) that cholera is one of the main causes of gastroenteritis in Northern Nigeria.

The common denominator in all the areas (Argungu, Jos, Suleja, Kwali and Gwagwa) is that they lack clean, portable water for consumption and generally rely on streams, ponds or wells that are exposed to sewage contamination. This situation spreads the disease rapidly.

The only limiting factor was that the survey was conducted in the dry season November 2010 – March 2011 when the communities had no rainfall and no overflowing streams that facilitates the spread of the disease.

Table 2 shows the sources of the samples and number collected in relation to the number of positive *Vibrio cholerae* isolated, Argungu with only 27 (17.3%) of the total samples collected had the highest number of positive samples which were 5 (18.5%), this is instructive showing that the number of samples collected does not determine the isolation of *Vibrio cholerae* but might be due to other factors which are reflected in other tables.

Table 3 depicts the age group of patients having gastroenteritis in relation to the location where the samples were collected. Most of the people that suffered from the outbreak based on the number of samples gotten were people from the age range of 20 – 29 (34.6%) and the least were people of the age range 50 – 59(1.9%). This could be explained with the fact that people at this age range (20- 29) are the most active, mobile and exposed of these age groups so their contact with contaminated food or water is likely to be more frequent thus reflecting in the number of cases gotten. All (100%) of the samples were Ogawa serotype which correlates with the findings of Linda et al, 2008 that the predominant serotype in this area (Sub-Saharan Africa) is Ogawa strain.

The Antibiotic sensitivity pattern of *Vibrio cholerae* isolated were as follows; all (100%) were sensitive to Ceftazidime, Cefuroxime Axetil, Gentamycin, Sparfloxacin and Tetracycline and all were resistant to Ampicillin, Cloxacillin, Streptomycin and Penicillin and thus is in consonance with the findings of Chander et al, 2008.

Most of the positive samples for *Vibrio cholerae* (7) were isolated from patients that had not been to the hospital or had spent less than 2 days in the hospital. It is evident that as the number of days spent in the hospital increased, the chances of isolating *Vibrio cholerae* from their stool samples decreased corresponding with the work of Umoh, *et al*, 2003.

Conclusion and recommendations
**Vibrio cholerae** is much feared because it is a killer disease; proper enlightenment campaign among the populace is of paramount importance.

It is not frequently sought for in bacteriology laboratories in most government hospitals. Government should assist in the control of cholera not only in the cities but also in the rural areas where people are more prone to infection.

During an epidemic, the critical elements of cholera control are early identification of cases through surveillance and case-finding, notification to health authorities and WHO, establishment of treatment centres, health education and proper disposal of human waste. Travellers to epidemic areas should be extremely careful about what they eat or drink and scrupulous about personal hygiene. Under the international health regulations, notification of WHO about cases of cholera is mandatory. National Health Authorities should report the first suspected cases to WHO at the earliest possible moment: laboratory confirmed cases should also be reported immediately. Thereafter, health authorities should report confirmed cases on a weekly basis.

National risk management strategies should be planned and implemented by a national coordinating committee. This committee should be responsible for cholera preparedness, intersectorial cooperation, regional and interregional collaboration, collection and reporting of information on cholera cases and deaths, organization of any special training that may be required, procurement, storage and distribution of essential supplies and implementation, supervision, monitoring and evaluation of control activities.

Through the course of this research it was found out that the microbiology laboratories of most of the government hospitals in the epidemic areas were not equipped with materials necessary for the isolation of *Vibrio cholera* even in epidemic periods.

However, since cholera does not give notice of its arrival, laboratories should be prepared. At least alkaline peptone water and Thiosulphate Citrate Bile-salt Sucrose Agar should be available.

Cholera is usually transmitted through the faecal-oral route, with the infective dose being around $10^8$. Individuals with reduced gastric acidity and blood group O are more susceptible to the infection. In situations where poor environmental sanitation, poor domestic and personal hygiene are rampant together with unavailability of clean, portable drinking water; transmission of cholera is more likely.

It has been suggested that control of cholera epidemics is too big a task for a national control programme, although this may be true to some extent in cases of extensive epidemics but a properly organized national control programme will provide the necessary framework for quick detection and prompt containment. This is also the best means for ensuring preparedness of cholera control.

Safe and effective cholera vaccines are available and constitute important tools for cholera control.

This study showed that *Vibrio cholerae* is one of the causes of gastroenteritis outbreak in Northern Nigeria.

References


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