

Molecular Docking of Selected Compounds Against Cellular Components of Bacteria

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Abstract

Researchers are trying to develop new antibiotics by targeting cellular components due to the emergence of antibiotic resistance by microbes. In this study, three targets were chosen these are penicillin binding protein-4, cell division protein, FtsA and shikimate dehydrogenase enzyme. Their binding sites were predicted online by RaptorX and GalaxyWEB servers. Virtual screening was carried out using the AutoDock Vina tool for a total of 50 experimental and approved compounds selected from the Drug Bank database. The results were redocked again by iGEMDOCK and the online SWISS-DOCK server. The top ten compounds in AutoDock Vina were selected. In Pharmacokinetics and pharmacodynamics study in silico, the highest three compounds in docking scores, Flunisolide, Doxazosin and Isradipine, showed high absorption by the gastrointestinal route and did not appear to cross blood blood-brain carrier, but the last two showed a probability of drug interaction via cytochrome. Hence, the study of pharmacokinetics and toxicities is crucial in the drug design approach. The use of more than one tool is preferred to obtain more reliable results.

Keywords: Docking, Molecular Targets, Pharmacokinetics, Virtual Screening.

Introduction

Bacteria are showing resistance to almost all currently used antibiotics via several mechanisms; therefore, researchers are investigating new alternative targets against which antibiotics may be developed, such as peptidoglycan synthesis, division machinery and shikimate biosynthetic pathway [1, 2].

Penicillin binding proteins (PBPs), sensitive to penicillin, are involved in peptidoglycan synthesis. These proteins catalyze polymerization of the glycan stand, which is composed of alternating N-acetylglucosamine and N-acetylmuramic acid through a transglycosylation reaction. Also, they are responsible for the cross-linking of the strands by transpeptidation. Some of these PBPs catalyze carboxypeptidation to bring about hydrolysis of the last amino acid of the pentapeptide and endopeptidation reaction,

which is the hydrolysis of the bond that connects two strands of glycan [3-5].

FtsA is another molecular target. FtsA is a component of the cell division machinery called the divisome. One of the important divisome proteins on the inner side of the plasma membrane is FtsZ, which forms bundles of protofilaments organised later into the Z-ring assembly. FtsA anchors FtsZ to the membrane. FtsA acts to interfere with the lateral interactions between FtsZ protofilaments and may affect FtsZ's high-order structure and the function of the divisome [6-8].

The shikimate acid pathway is involved in aromatic amino acids synthesis in bacteria, fungi and plants. It is absent in humans, making its enzyme components a good target to design antibiotics. Shikimate dehydrogenase is responsible for the

conversion of 3-dehydroshikimate acid into shikimate using NADPH [1]. In this study, virtual screening is performed to identify ligands from the Drug Bank database to inhibit three possible targets, namely PBP-4, FtsA and shikimate dehydrogenase, through a molecular docking approach.

Materials and Methods

Obtaining protein models and prediction of binding sites: Crystal structures of three targets were obtained from the Protein Data Bank database. The crystal structure of PBP-4 (dacB) has the PDB ID: 2EX8, the crystal structure of FtsA has PDB ID: 3WQU, and that of shikimate dehydrogenase has PDB ID: 1NYT. Binding sites were predicted by the RaptorX online server developed by Källberg *et al.* [9] at (<http://raptorx.uchicago.edu/>). GalaxyWEB web server [10] was also available at: (<http://galaxy.seoklab.org/cgi-bin/submit.cgi?type=SITE>).

Molecular docking: A total of 50 experimental and approved compounds from the Drug Bank database were selected [11]. It also shows the molecular descriptors of compounds. The controls were Penicillin G, ATP and shikimate for PBP-4, FtsA and shikimate dehydrogenase, respectively. Molecular docking was performed by three docking tools. AutoDock Vina [12] used an autogrid tool to pre-calculate a grid. This grid has a size of 60×60 × 60 and a box centre of 87.709, 4.765 and 45.407 for x, y and z, respectively, for the PBP-4 target. A box centre at -2.825, 25.163 and -32.295 for FtsA target was used and 57.345, 27.835 and 20.36 for shikimate dehydrogenase. Interactions between compounds and target models were visualized by LIGPLOT+ [13]. Results were subjected to docking with iGEMDOCK [14]. The online SWISS-DOCK was employed to dock the results again [15]. It is available at <http://www.swissdock.ch/>.

Pharmacokinetics and pharmacodynamics predictions: SwissADME online program was used to predict the pharmacokinetics and pharmacodynamics of the experimental and approved compounds [16]. It can be accessed at: <http://www.swissadme.ch/>.

Results and Discussion

Three docking tools were used in the virtual screening of 50 experimental and approved chemical compounds obtained from the Drug Bank database, and the top ten compounds in docking scores were presented according to Autodock Vina. These ten compounds were re-docked twice by different tools to get more accurate results. The interaction between molecular targets and these ten ligands in terms of hydrogen bonds and hydrophobic interactions.

PBPs are classified into two main groups. High molecular weight PBPs are subdivided into Class A and B. These act in glycosylation and transpeptidation reactions involved in polymerisation of the peptidoglycan sacculus and its insertion into the preformed cell wall [17, 18]. There are seven low molecular weight PBPs, referred to as Class C, such as PBP4 and PBP7. These are endopeptidases that can cleave cross-linking between two strands of glycan, as in recycling, cell separation and peptidoglycan maturation [19].

The crystal structure of PBP-4 (dac) was used in this study [20]. PBP-4 consists of three domains. Two domains are inserted in the transpeptidase domain in the way of “matryoshka dolls”, where the third domain is inserted in the second domain, which is inserted in the penicillin-binding domain [4]. RaptorX predicted the binding residues as follows: A⁴², S⁴³, K⁴⁶, F¹⁴¹, S²⁸⁷, N²⁸⁹, G³³⁹, L³⁴⁰, K³⁹⁸, T³⁹⁹ while GalaxyWEB server: A⁴², S⁴³, S²⁸⁷, N²⁸⁹, S³⁸⁸, G³³⁹, L³⁴⁰, G⁴⁰⁰, S⁴⁰¹, L⁴⁰² (Table 1 and Table 2).

Table 1. Docking Scores (Kcal/mol) of PBP-4

| Compound | AutoDock Vina | iGEMDOCK | SWISS-DOCK |
|----------|---------------|----------|------------|
| PNG | -6.2 | -54.28 | -7.8 |
| DB00177 | -7.1 | -82.7 | -7.7 |
| DB00180 | -7.5 | -74.6 | -7.3 |
| DB00243 | -7.2 | -76.5 | -7.2 |
| DB00270 | -7.2 | -75.4 | 7.9 |
| DB00276 | -7.3 | -75.0 | -7.4 |
| DB00301 | -7.2 | -71.2 | 7.8 |
| DB00443 | -7.1 | -73.1 | -7.6 |
| DB00485 | -7.0 | -67.5 | -7.7 |
| DB00494 | -7.0 | -79.8 | -8.0 |
| DB00522 | -7.4 | -77.3 | -7.9 |

Table 2. Interaction of PBP-4 with Ligands

| Compound | Hydrogen bonds | Hydrophobic interactions |
|----------|------------------|---|
| PNG | S398 | S62, F160, K305, S306, N308, L359, T418, G419, S420, L421 |
| DB00177 | N308 | C159, F160, R171, K305, S306, D307, L359, S420, Q422 |
| DB00180 | S62, S240, S306 | F160, R171, N308, L421 |
| DB00243 | - | F160, R171, K305, S306, D307, L359, S420, L421 |
| DB00270 | - | S62, F160, S306, N308, L359, S420, L421, Q422 |
| DB00276 | - | S62, F160, S306, D307, N308, R361, S420, L421, Q422 |
| DB00301 | - | S62, F160, R171, S306, D307, N308, L359, L421 |
| DB00443 | S306, T418, S420 | S62, F160, K305, G419 |
| DB00485 | - | S62, F160, R171, S306, D307, N308, L359, L421 |
| DB00494 | - | S62, F160, R171, S306, D307, N308, L359, L421 |
| DB00522 | R171, R361, Q422 | C159, F160, N308, L359, L421 |

The FtsA crystal structure in complex with an ATP molecule was used [21]. FtsA resembles actin in its structure and appears to consist of two domains with a common core, which makes a groove between these domains where the nucleotide binds. Each domain consists of two subdomains; A1 and A2 are large and composed of five β -sheets and three α -helices, while the other two are variable in

the actin family [22]. RaptorX predicted the following amino acid residues in binding site: G²⁸, S²⁹, S³⁰, S³¹, K³³, G²²⁴, E²²⁵, D²²⁶, V²²⁷, G²⁴⁸, E²⁶⁷, K²⁷⁰, H²⁷¹, G³⁴⁰, G³⁴¹, S³⁴², N³⁴⁴, L³⁴⁵, E³⁷⁴ while GalaxyWEB server predicted binding site as follows: G²⁸, S²⁹, S³¹, K³³, G²²⁴, E²²⁵, D²²⁶, V²²⁷, K²⁷⁰, H²⁷¹, G³⁴⁰, G³⁴¹, S³⁴², N³⁴⁴, L³⁴⁵, E³⁷⁴ (Table 3 and Table 4).

Table 3. Docking Scores (Kcal/mol) of FtsA

| Compound | AutoDock Vina | iGEMDOCK | SWISS-DOCK |
|----------|---------------|----------|------------|
| ATP | -8.8 | -61.3 | -8.14 |
| DB00197 | -9.1 | -61.2 | -7.9 |
| DB00243 | -9.6 | -78.8 | -7.2 |

| | | | |
|---------|-------|-------|------|
| DB00301 | -8.3 | -69.4 | -7.4 |
| DB00341 | -8.9 | -85.3 | -7.6 |
| DB00346 | -8.7 | -70.1 | -8.3 |
| DB00384 | -9.1 | -93.7 | -7.5 |
| DB00436 | -9.2 | -86.1 | -7.3 |
| DB00469 | -8.5 | -96.4 | -7.9 |
| DB00507 | -10.1 | -87.8 | -8.5 |
| DB00590 | -10.9 | -79.9 | -7.6 |

Table 4. Interaction of FtsA with Ligands

| Compound | Hydrogen bonds | Hydrophobic interactions |
|----------|---------------------------|---|
| ATP | S15 | G12, S14, P79, G208, E209, V211, Q213, H225, K254, G325, S326, N328, L329 |
| DB00197 | D185 | D10, G12, S14, K17, K77, T189, D206, G208, E209, V211, K254, H255, G325, S326, N328, E358, S361 |
| DB00243 | - | D10, G12, S13, S14, K17, K77, D206, G208, E209, V211, Q213, G325, S326, N328, G232, H255, E358 |
| DB00301 | D10, S14, E209, G325 | G12, S15, K17, Q35, D206, G208, G324, S236, K254, H255, E358 |
| DB00341 | S14, E358 | D10, G12, S13, S15, K17, Y37, D206, G208, E209, V211, G324, G325, E358, S361 |
| DB00346 | S15, K17, K77, D206, H255 | D10, G12, S13, S14, G44, G208, V211, Q213, K254, G325, S326, N328, E358 |
| DB00384 | - | D10, G12, S14, K17, K77, D206, G208, E209, Q213, G324, G325, E358, S361 |
| DB00436 | S13, K17, K77, D206, G325 | S14, P79, G208, E209, V211, Q213, G324, E358 |
| DB00469 | K17, E358 | D10, G12, S13, S14, K77, D185, W189, D206, G208, E209, V211, Q213, G325, S361 |
| DB00507 | K77 | S13, S14, K17, D206, G208, E209, V211, E213, G325, E358, S361 |
| DB00590 | - | D10, S13, S14, S15, K17, Q35, D206, G208, E209, Q213, K254, H255, G325, S326, N328, E358, S361 |

The Shikimate dehydrogenase crystal structure solved by Michel *et al.* [23] was used. Shikimate dehydrogenase contains two domains. The first catalytic domain possesses a twisted α/β motif. The second domain is a Rossmann fold in configuration and binds to

NADPH [24]. According to RaptorX, the following residues are present in the binding site: V⁶, S¹⁴, S¹⁶, N⁵⁹, V⁶⁰, T⁶¹, K⁶⁵, N⁸⁶, D¹⁰², L²⁴¹, Q²⁴⁴ while GalaxyWEB server shows the following: S¹⁶, N⁵⁹, T⁶¹, K⁶⁵, N⁸⁶, D¹⁰², L²⁴¹, Q²⁴⁴ (Table 5 and Table 6).

Table 5. Docking Scores (Kcal/mol) of Shikimate Dehydrogenase

| Compound | AutoDock Vina | iGEMDOCK | SWISS-DOCK |
|-----------|---------------|----------|------------|
| Shikimate | -6.5 | -62.8 | -6.4 |
| DB00177 | -9.2 | -95.8 | -9.3 |
| DB00197 | -9.8 | -76.8 | -8.5 |
| DB00203 | -9.2 | -88.1 | -8.9 |

| | | | |
|---------|-------|-------|------|
| DB00243 | -9.4 | -84.0 | -8.3 |
| DB00270 | -10.3 | -86.3 | -8.6 |
| DB00276 | -9.4 | -88.7 | -8.9 |
| DB00401 | -9.8 | -79.2 | -8.5 |
| DB00507 | -9.3 | -89.6 | -7.8 |
| DB00522 | -10.0 | -90.8 | -8.2 |
| DB00590 | -9.5 | -72.9 | -8.8 |

Table 6. Interaction of Shikimate Dehydrogenase with Ligands

| Compound | Hydrogen bonds | Hydrophobic interactions |
|-----------|--------------------------|---|
| Shikimate | S16, V60, T61, N86, D102 | D59, K65, L241, Q244 |
| DB00177 | D102 | V6, S16, N59, V60, T61, V62, K65, A187, T188, S189, M213, F214, W215, L241, Q244 |
| DB00197 | S16, N86, K65 | V6, P10, N59, V60, T61, V62, D102, T188, S189, I192, M213, L241, Q244 |
| DB00203 | S14 | V6, P10, S16, N59, T61, V62, D102, A130, A187, T188, S189, M213, F214, W215, L241, Q244 |
| DB00243 | - | V6, S14, S16, V60, T61, N86, A130, T188, S189, M213, F214, W215, L241, Q244 |
| DB00270 | - | T61, V62, K65, A130, A187, S189, M213, W215 |
| DB00276 | K65, A187 | V62, P63, D102, G128, G129, A130, T188, S189, M213 |
| DB00401 | T61 | P10, V62, D102, T188, S189, I192, M213, F214, W215, M240, L241 |
| DB00507 | S16, Q244 | V6, S14, N59, T61, W215, L241 |
| DB00522 | W215 | V62, P63, K65, G129, A130, A187, T188, S189, M213, F214 |
| DB00590 | - | S14, T61, V62, P63, K65, E66, G128, G129, A187, T188, S189, M213, F214, W215, L241 |

Flunisolide (DB00180) had the highest docking score in the virtual screening targeting PBP-4. It is a synthetic corticosteroid used in

the treatment of asthma as an inhaler [25] (Table 7 and Figure 1).

Table 7. Molecular Descriptors of the Tested Compounds

| Compound | Name | Molecular weight | logP | Hydrogen bond donors | Hydrogen bond acceptor |
|----------|----------------|------------------|------|----------------------|------------------------|
| DB00177 | Valsartan | 435.519 | 3.68 | 2 | 6 |
| DB00180 | Flunisolide | 434.498 | 2.20 | 2 | 6 |
| DB00197 | Troglitazone | 441.540 | 4.16 | 2 | 5 |
| DB00203 | Revatio | 474.576 | 2.35 | 1 | 8 |
| DB00243 | Ranolazine | 427.536 | 2.08 | 2 | 6 |
| DB00270 | Isradipine | 371.387 | 3.00 | 1 | 5 |
| DB00276 | Amsacrine | 393.459 | 4.66 | 1 | 5 |
| DB00301 | Flucloxacillin | 453.056 | 2.69 | 2 | 5 |
| DB00341 | Cetirizine | 388.888 | 2.98 | 1 | 5 |

| | | | | | |
|---------|---------------------|---------|------|---|---|
| DB00346 | Alfuzosin | 389.449 | 2.02 | 2 | 8 |
| DB00384 | Triamterene | 253.263 | 1.21 | 3 | 7 |
| DB00401 | Nisoldipine | 388.414 | 3.63 | 1 | 5 |
| DB00436 | Bendroflumethiazide | 421.415 | 1.83 | 3 | 5 |
| DB00443 | Betamethasone | 392.461 | 1.93 | 3 | 5 |
| DB00469 | Tenoxicam | 337.370 | 2.42 | 2 | 5 |
| DB00485 | Dicloxacillin | 470.326 | 3.19 | 2 | 5 |
| DB00494 | Entacapone | 305.286 | 2.50 | 2 | 6 |
| DB00507 | Nitazoxanide | 307.282 | 2.14 | 1 | 5 |
| DB00522 | Bentiromide | 404.415 | 2.99 | 4 | 5 |
| DB00590 | Doxazosin | 451.475 | 2.53 | 5 | 9 |

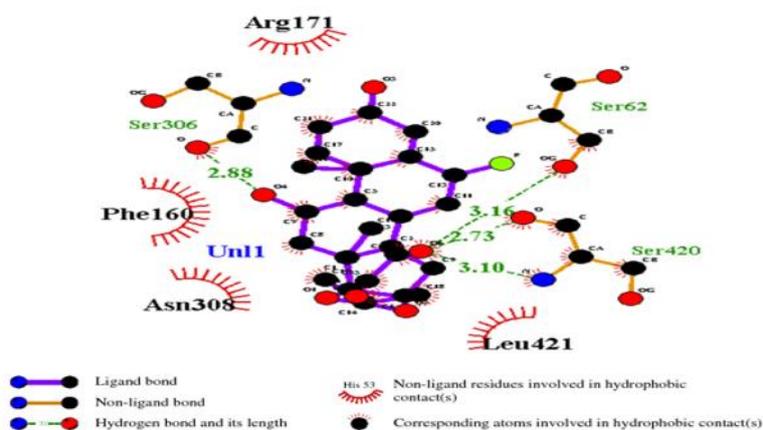


Figure 1. Interaction of PBP with DB00180

Doxazosin (DB00590) showed the highest docking score in AutoDock in the case of the FtsA target. It is an alpha1 blocker drug used in the treatment of benign prostatic hyperplasia [26]. Isradipine (DB00270), a

calcium channel blocker used in the treatment of hypertension [27], showed the highest docking score against shikimate dehydrogenase. Molecular descriptors (Figure 2).

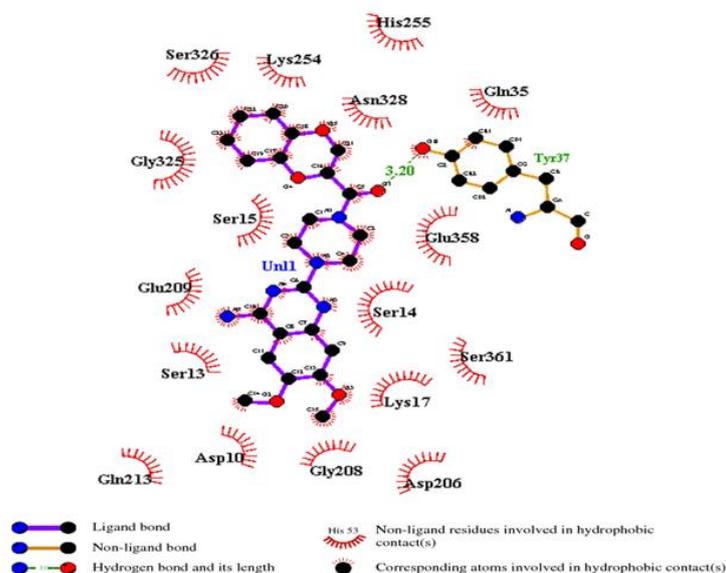


Figure 2. Interaction of FtsA with DB00590

José Alves et al. [28] used AutoDock 4 to measure the inhibition constant (Ki) of compounds found in mushrooms against PBP1a. Vanillic acid, gallic acid and protocatechuic acid have Ki values of 19.05, 19.02 and 18.2 μm , respectively. Kulanthaivel et al. [29] found that a compound which had ZINC database ID 95911396 gave a docking score of -10.12 among others, which was higher than previously reported ligands (3-

bromopyruvate, 2-deoxyglucose, lonidamine, imatinib and oxythiamine). In addition, Isa et al. [30] used Autodock4 in virtual screening of a total of 13803 compounds against shikimate dehydrogenase (Figure 3). The study identified 26 compounds with binding energies ranged between -12.03 to -8.33 Kcal/mol. In further analyses, two compounds namely ZINC12135132 and ZINC08951370 were identified to have the best inhibitory actions.

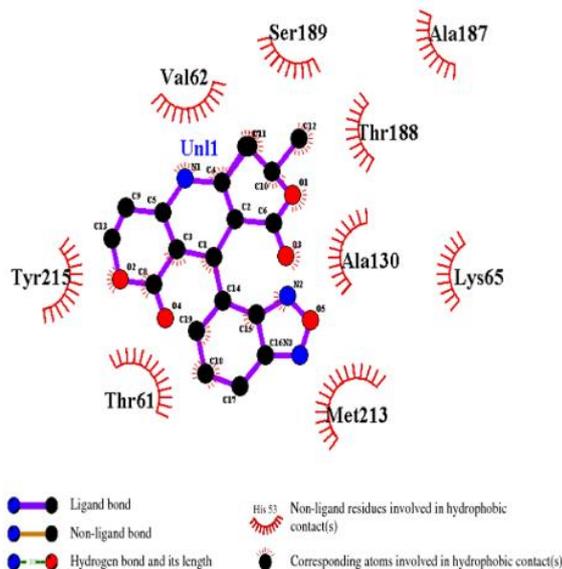


Figure 3. Interaction of Shikimate Dehydrogenase with DB00270

Table 8 shows some predictions of pharmacokinetics and pharmacodynamics for the experimental and approved compounds. Flunisolide, Doxazosin and Isradipine have high gastrointestinal absorption and do not penetrate blood blood-brain barrier, so they can enter the body easily and do not have toxicity for the central nervous system [31]. However, both Doxazosin and Isradipine may

inhibit cytochromes that are responsible for the oxidative stage of drug elimination from the human body [32], implying that toxicity from other drugs may result, i.e. drug interaction [33]. This green chemistry provides a valuable technique to control infection [34], even in the presence of multidrug-resistant bacteria [35] or associated chronic diseases [36].

Table 8. Pharmacokinetics and Pharmacodynamics of Tested Compounds

| Compound | G.I. | BBB | P-gp | Cytochrome inhibition | | | | | D.L. |
|----------|------|-----|------|-----------------------|------|-----|-----|-----|------|
| | | | | 1A2 | 2C19 | 2C9 | 2D6 | 3A4 | |
| DB00177 | High | No | No | No | Yes | Yes | No | Yes | Yes |
| DB00180 | High | No | Yes | No | No | No | No | No | Yes |
| DB00197 | High | No | Yes | No | Yes | Yes | No | Yes | Yes |
| DB00203 | High | No | Yes | No | No | Yes | No | Yes | Yes |
| DB00243 | High | No | Yes | No | No | No | Yes | No | Yes |
| DB00270 | High | No | No | Yes | Yes | Yes | No | Yes | Yes |
| DB00276 | High | No | No | Yes | Yes | Yes | Yes | Yes | Yes |

| | | | | | | | | | |
|---|------|-----|-----|-----|-----|-----|-----|-----|-----|
| DB00301 | Low | No | Yes | No | Yes | No | No | Yes | Yes |
| DB00341 | High | Yes | Yes | No | No | No | Yes | No | Yes |
| DB00346 | High | No | Yes | No | No | No | No | No | Yes |
| DB00384 | High | No | Yes | Yes | No | No | Yes | Yes | Yes |
| DB00401 | High | No | No | Yes | Yes | Yes | Yes | Yes | Yes |
| DB00436 | Low | No | Yes | No | No | No | No | No | Yes |
| DB00443 | High | No | Yes | No | No | No | No | No | Yes |
| DB00469 | High | No | Yes |
| DB00485 | Low | No | Yes | No | Yes | No | No | Yes | Yes |
| DB00494 | Low | No | Yes | No | Yes | No | No | Yes | Yes |
| DB00507 | Low | No | No | No | Yes | No | No | No | Yes |
| DB00522 | High | No | No | No | No | Yes | Yes | No | Yes |
| DB00590 | High | No | No | No | No | Yes | Yes | Yes | Yes |
| G.I.: Gastrointestinal absorption, BBB: Blood brain barrier penetration, P-gp: Plasma glycoprotein substrate, D.L.: Lipinski's drug likeness. | | | | | | | | | |

Conclusion

The objective of the virtual screening approach is to select compounds for further *in vitro* experiments, where thousands of compounds are subjected to molecular docking against a given target and the top ten compounds are scored. However, negative and positive false results may occur. Using more than one docking tool can improve the process of getting more reliable results.

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Conflict of Interest

The authors declare no conflict of interest.

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