

Molecular Docking of Phytochemicals from *Adhatoda vasica* Against Caries, Periodontitis and Inflammatory Mediators – A Computational Study

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Abstract

Orthodontic treatment is associated with pain, difficulty in plaque control, caries incidence, and slow movement of teeth. Authors hypothesize that the answer to these questions may lie in phytochemicals which are a huge unexplored area. Due to previous reports on *Adhatoda vasica* about its anti-inflammatory and antimicrobial properties, its use in orthodontics is explored in the dry lab scenario. This can be considered as a preliminary step to study the herb in great detail for use in orthodontic therapy. Computational algorithms from Auto Dock version 4 were used in the study. Vasicine, Vasinone, Vasicoline and Anisotine were analysed against Glucosyltransferase (GFT) of *Streptococcus mutans*, Gingipain K - *Porphyromonas gingivalis*, FIM A of *Porphyromonas gingivalis*, TNF ALPHA and Prostaglandin H synthases. Compounds exerted promising inhibiting action against *Streptococcus mutans*, Gingipain K, FimA, TNF-alpha and prostaglandin E2. The said phytochemicals - Vasicine, Vasicinone, Vasicoline and Anisotine can further be explored for proper delivery to saliva, periodontal region and to bone for effective use during and after orthodontic treatment.

Keywords: *Adhatoda vasica*, docking, Glucosyltransferase, Gingipain K - *Porphyromonas gingivalis*, FIM A of *Porphyromonas gingivalis*, TNF ALPHA and Prostaglandin H synthases.

Introduction

In nature, caries and periodontitis do not commonly exist together, except in cases of very poor oral hygiene [1]. This is because distinct flora are associated with caries and periodontitis which antagonize each other [2]. However, rare situations like orthodontic treatment cause an increase in both cariogenic and periodontal flora. As mentioned, orthodontic treatment, especially using fixed appliances greatly hinders the maintenance of oral hygiene. Therefore, various aids are used to promote this cleaning process to prevent caries and periodontitis, which are equally deleterious to the patient [3]

agent that can achieve all this. For controlling pain, NSAIDs are used and duration may not

Of various modalities used to treat the situation, after physical techniques of brushing, chemical methods come into play. Mouthwashes play a major role in plaque control in these situations. Though they are successful, numerous adverse events like damaging mucosa, staining of teeth, and altering taste perception are associated with it [4].

In another direction, many patients refuse orthodontic treatment due to the pain associated with it and its long duration [5]. If these issues can be addressed, acceptance of orthodontic treatment would increase and more patients would benefit from it. In the present scenario, there is no single be effectively controlled by drugs or chemicals [6].

Authors hypothesize that the answer to these questions may lie in phytochemicals which are a huge unexplored area. In that direction, due to previous reports on *Adhathoda vasica* about its anti-inflammatory and antimicrobial properties, its use in orthodontics is explored in the dry lab scenario. This can be considered as a preliminary step to study the herb in great detail for use in orthodontic therapy.

Materials and Methods

Protein-Ligand Docking

Computational algorithms predicting the possibility of leads over selected targets were performed by using a standard docking tool (Auto Dock version 4). The outcome of screening was ascertained by observing the binding affinity and interaction pattern of the lead with targets [7].

Protein Preparation

The three-dimensional structure of the target proteins illustrated in Figure 1 was retrieved from the Research Collaboratory for Structural Bioinformatics (RCSB). The retrieved protein construct was subjected to surface optimization by removing the preloaded lead candidate and subsequent water molecules cleaved. Gasteiger charges were computed with additional polar hydrogen atoms, and the merging of non-polar and rotatable bonds was defined using AutoDock 4 [8].

Ligand Model Preparation

Two- and three-dimensional structures of selected ligands viz. Vasicine, Vasicinone, Vasicoline and Anisotine subjected to molecular docking investigation were constructed by using ChemDraw sketch software. Figure 2 represents the 2D and 3D structures of selected ligand molecules subjected to molecular docking investigation.

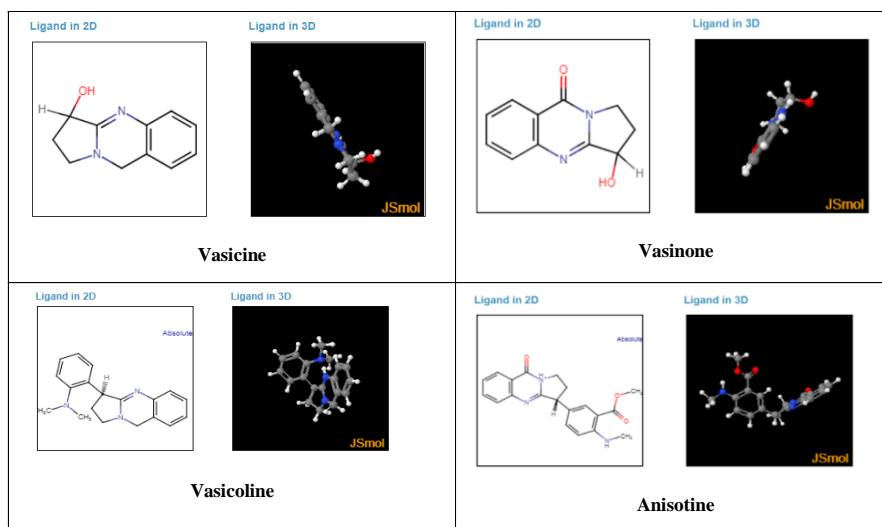


Fig. 1. 2D and 3D Structure of Phytocomponents

Table 1. Ligand Properties of the Compounds Selected for Docking Analysis

| Compound | Molar Weight g/mol | Molecular Formula | H Bond Donor | H Bond Acceptor | Rotatable Bonds |
|------------|--------------------|---|--------------|-----------------|-----------------|
| Vasicine | 188.23 g/mol | C ₁₁ H ₁₂ N ₂ O | 1 | 2 | 0 |
| Vasicinone | 202.213 g/mol | C ₁₁ H ₁₀ N ₂ O ₂ | 1 | 3 | 0 |
| Vasicoline | 291.4 g/mol | C ₁₉ H ₂₁ N ₃ | 0 | 2 | 2 |
| Anisotine | 349.4 g/mol | C ₂₀ H ₁₉ N ₃ O ₃ | 1 | 5 | 4 |

Docking Simulations

In-silico docking predictions were performed using a licensed version of AutoDock 4. The virtual screening tool works behind the algorithm to predict the possible interaction between the functional group of the lead with the active site (amino acid residue) of the protein target. The three-dimensional conformation of target proteins retrieved from RCSB was subjected to computational docking by using AutoDock 4. Molecular energy and free dynamics calculation were optimized using the auto dock algorithm and protein charge calculation set with the Gasteiger simulation module. Solvation parameters and other respective polar hydrogens were added to the receptor for the preparation of protein for successive docking

simulation. Since ligands are phytotherapeutics and not of peptide origin hence, Gasteiger charges were essentially applied and then non-polar hydrogens were merged. As per the requirement of the AutoDock programming, the pre-calculated grid maps were set for one for each atom type, present in the ligand being docked as it stores the potential energy arising. The docking pocket was set with affinity (grid) maps of 70×70×70 Å grid points and with 0.375 Å. Each docking calculation was derived from 10 different runs that were set to terminate after a maximum of 250000 energy evaluations. The population size was set to 150. During the search, a translational step of 0.2 Å, and quaternion and torsion steps of 5 were applied [9, 10].

Table 2. Details of Targets

| S. No. | Purpose | PDB | Name of Target |
|--------|--|-----------|--|
| 1 | Anti-Microbial activity - Caries | 3AIC [11] | Glucosyltransferase (GFT) of <i>Streptococcus mutans</i> |
| 2 | Anti-microbial activity - Periodontal | 6I9A | Gingipain K - <i>Porphyromonas gingivalis</i> |
| 3 | Anti-microbial activity - Periodontal | 4Q98 | FIM A of <i>Porphyromonas gingivalis</i> |
| 4 | Analgesic / Anti-Inflammatory Activity | 2AZ5 | TNF ALPHA |
| 5 | Anti-Inflammatory Activity | 1IGX | Prostaglandin H synthases |

Receptor Structures

The crystalline structure of the target proteins was retrieved from the protein data bank and protein clean-up process was done and essential missing hydrogen atoms were added. Different orientation of the lead molecules concerning the target protein was evaluated by the Autodock program and the best dock pose was selected based on the interaction study analysis.

Docking calculations were carried out for retrieved phytocomponents against target protein 3CL pro. Essential hydrogen atoms, Kollman united atom type charges, and solvation parameters were added with the aid of AutoDock tools.[10] Affinity (grid) maps of $\times\times$ Å grid points and 0.375 Å spacing were generated using the Autogrid program. AutoDock parameter set- and distance-dependent dielectric functions were used in the calculation of the van der Waals and the electrostatic terms, respectively. Docking simulations were performed using the Lamarckian genetic algorithm (LGA) and the Solis & Wets local search method [12]. The initial position, orientation, and torsions of the ligand molecules were set randomly. All rotatable torsions were released during

docking. Each docking experiment was derived from 2 different runs that were set to terminate after a maximum of 250000 energy evaluations. The population size was set at 150. During the search, a translational step of 0.2 Å, and quaternion and torsion steps of 5 were applied [13, 14, 15].

In-silico docking simulations were performed by using Auto Dock version 4. The molecular interaction of residual amino acids with the core functional groups determines the efficacy of the lead molecules. Three dimensional pharmacophores of FDA-approved lead molecules were subjected to virtual screening against selected protein target Gingipain K - *Porphyromonas gingivalis* with PDB 6I9A retrieved from RCSB by using Auto Dock 4. Docking grids were set with the pocket size measuring maps of 70×70×70 Å grid points and with 0.375 Å. Each docking calculation is set to run with 10 different cycles after a maximum of 250000 energy evaluations. The population size was set at 150. During the search, a translational step of 0.2 Å, and quaternion and torsion steps of 5 were applied [7, 16].

Results

Table 3. Summary of the Molecular Docking Studies of Compounds Against Glucosyltransferase (GFT) of *Streptococcus mutans*- PDB- 3AIC

| Compounds | Binding Free Energy Kcal/mol | Inhibition Constant Ki μ M (*mM)(**nM) | Electrostatic Energy Kcal/mol | Intermolecular Energy Kcal/mol | Total Interaction Surface |
|------------|------------------------------|--|-------------------------------|--------------------------------|---------------------------|
| Vasicine | -7.02 kcal/mol | 7.20 μ M | -2.79 kcal/mol | -7.31 kcal/mol | 539.729 |
| Vasicinone | -5.01 kcal/mol | 212.78 μ M | -0.07 kcal/mol | -5.31 kcal/mol | 464.886 |
| Vasicoline | -9.24 kcal/mol | 169.65 nM | -2.70 kcal/mol | -9.42 kcal/mol | 671.964 |
| Anisotine | -7.04 kcal/mol | 6.94 μ M | -0.01 kcal/mol | -7.55 kcal/mol | 662.994 |

Table 4. Amino acid Residue Interaction of Lead against Glucosyltransferase (GFT) of *Streptococcus mutans*-PDB- 3AIC

| Compounds | Interaction | Amino Acid Residues | | | | | | | | | | |
|------------|-------------|---------------------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|
| | | 433 LEU | 475 ARG | 477 ASP | 478 ALA | 481 ASN | 515 TRP | 517 TRP | 587 HIS | 588 ASP | 916 TYR | |
| Vasicine | 3 | 433 LEU | 475 ARG | 477 ASP | 478 ALA | 481 ASN | 515 TRP | 517 TRP | 587 HIS | 588 ASP | 916 TYR | |
| Vasicinone | 3 | 433 LEU | 477 ASP | 478 ALA | 480 ASP | 481 ASN | 515 TRP | 517 TRP | 588 ASP | | | |
| Vasicoline | 3 | 430 TYR | 433 LEU | 475 ARG | 477 ASP | 480 ASP | 481 ASN | 515 TRP | 517 TRP | 587 HIS | 588 ASP | 916 TYR |
| Anisotine | 3 | 433 LEU | 477 ASP | 515 TRP | 517 TRP | 588 ASP | 589 SER | 592 GLN | 593 ASP | 610 TYR | 907 PHE | 916 TYR |

Table 5. Summary of the Molecular Docking Studies of Compounds Against Gingipain K - *Porphyromonas gingivalis* – PDB 6I9A

| Compounds | Binding Free Energy Kcal/mol | Inhibition Constant Ki μM (*mM)(**nM) | Electrostatic Energy Kcal/mol | Intermolecular Energy Kcal/mol | Total Interaction Surface |
|------------|------------------------------|--|-------------------------------|--------------------------------|---------------------------|
| Vasicine | -5.94 kcal/mol | 44.19 μM | -0.80 kcal/mol | -6.24 kcal/mol | 447.239 |
| Vasicinone | -5.93 kcal/mol | 45.06 μM | -0.10 kcal/mol | -6.23 kcal/mol | 424.733 |
| Vasicoline | -7.49 kcal/mol | 3.26 μM | -0.58 kcal/mol | -7.23 kcal/mol | 577.76 |
| Anisotine | -5.65 kcal/mol | 72.15 μM | -0.03 kcal/mol | -6.32 kcal/mol | 592.433 |

Table 6. Amino Acid Residue Interaction of Leads Against Gingipain K - *Porphyromonas gingivalis* – PDB 6I9A

| Compounds | Interaction | Amino acid Residues | | | | | | |
|------------|-------------|---------------------|---------|---------|---------|---------|---------|---------|
| | | 442 THR | 444 HIS | 477 CYS | 511 SER | 512 TYR | 513 TRP | 516 ASP |
| Vasicine | 2 | 442 THR | 444 HIS | 477 CYS | 511 SER | 512 TYR | 513 TRP | 516 ASP |
| Vasicinone | 2 | 444 HIS | 476 CYS | 477 CYS | 511 SER | 513 TRP | 516 ASP | |
| Vasicoline | 3 | 388 ASP | 391 TRP | 444 HIS | 477 CYS | 511 SER | 513 TRP | |
| Anisotine | 3 | 388 ASP | 390 SER | 391 TRP | 444 HIS | 477 CYS | 511 SER | 513 TRP |

Table 7. Docking Studies of Compounds Against FIMA – PDB 4Q98

| Compounds | Binding Free Energy Kcal/mol | Inhibition Constant Ki μM (*mM)(**nM) | Electrostatic Energy Kcal/mol | Intermolecular Energy Kcal/mol | Total Interaction Surface |
|------------|------------------------------|--|-------------------------------|--------------------------------|---------------------------|
| Vasicine | -5.37 kcal/mol | 115.99 μM | -0.04 kcal/mol | -5.67 kcal/mol | 521.66 |
| Vasicinone | -6.69 kcal/mol | 12.41 μM | -0.15 kcal/mol | -6.99 kcal/mol | 519.072 |
| Vasicoline | -7.54 kcal/mol | 2.95 μM | -0.01 kcal/mol | -8.04 kcal/mol | 664.194 |
| Anisotine | -5.25 kcal/mol | 141.27 μM | -0.05 kcal/mol | -5.38 kcal/mol | 641.019 |

Table 8. Amino Acid Residue Interaction of Leads Against FIMA with PDB 4Q98

| Compounds | Interaction | Amino Acid Residues | | | | | | | | | | |
|------------|-------------|---------------------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|
| | | | | | | | | | | | | |
| Vasicine | 1 | 237 TYR | 317 PRO | 319 LEU | 325 ASN | 363 PRO | 364 GLN | 367 GLN | | | | |
| Vasicinone | 1 | 237 TYR | 317 PRO | 319 LEU | 325 ASN | 327 TYR | 363 PRO | 364 GLN | 367 GLN | 368 ALA | 370 LEU | |
| Vasicoline | 1 | 237 TYR | 317 PRO | 319 LEU | 325 ASN | 327 TYR | 363 PRO | 364 GLN | 367 GLN | 368 ALA | | |
| Anisotine | 3 | 237 TYR | 317 PRO | 319 LEU | 325 ASN | 357 THR | 360 PRO | 363 PRO | 364 GLN | 367 GLN | 368 ALA | 370 LEU |

Table 9. Summary of the Molecular Docking Studies of Compounds Against TNF-alpha (2AZ5)

| Compounds | Binding Free Energy Kcal/mol | Inhibition Constant Ki μM (*mM)(**nM) | Electrostatic Energy Kcal/mol | Intermolecular energy Kcal/mol | Total Interaction Surface |
|------------|------------------------------|--|-------------------------------|--------------------------------|---------------------------|
| Vasicine | -4.41 kcal/mol | 581.81 μM | -0.19 kcal/mol | -4.71 kcal/mol | 378.73 |
| Vasicinone | -4.67 kcal/mol | 376.54 μM | -0.07 kcal/mol | -4.97 kcal/mol | 380.683 |
| Vasicoline | -6.21 kcal/mol | 27.83 μM | -0.04 kcal/mol | -5.89 kcal/mol | 472.27 |
| Anisotine | -6.25 kcal/mol | 26.10 μM | -0.11 kcal/mol | -6.87 kcal/mol | 606.919 |

Table 10. Amino Acid Residue Interaction of Lead Against TNF-alpha (2AZ5)

| Compounds | Interaction | Amino Acid Residues | | | | | |
|------------|-------------|---------------------|-----------|------------|------------|------------|------------|
| | | 59 | 61 | 119 | 151 | | |
| Vasicine | 3 | TYR | GLN | TYR | TYR | | |
| Vasicinone | 4 | 57 LEU | 59 TYR | 61 GLN | 119 TYR | 120 LEU | 151 TYR |
| Vasicoline | 4 | 57 LEU | 59 TYR | 119 TYR | 151 TYR | 155 ILE | |
| Anisotine | 4 | 57 LEU | 59 TYR | 61 GLN | 119 TYR | 151 TYR | |

Discussion

With regard to computational analysis related to the use of AV in orthodontics, five targets were chosen. Primarily, the major complication is the dental caries as sequelae of orthodontic treatment. Caries are caused by multiple groups of microbes, of which *Streptococcus mutans* is the predominant pathogen. Glycosyltransferase (Gtf) is a critical virulence factor of *Streptococcus mutans*. Extracellular polysaccharides, especially glucans produced by *S. mutans* Gtfs, appear to contribute to the cariogenicity of dental biofilms, according to all available evidence. As a result, inhibiting Gtf activity and the resulting polysaccharide synthesis may reduce the virulence of cariogenic biofilms, which could be an alternative strategy for disease prevention caused by biofilms [17]. In this study binding of the four phytochemicals/leads with the said molecule was analysed.

It was observed that all 4 leads such as Vasicine, Vasicinone, Vasicoline and Anisotine reveal maximum of 3 interactions with the core active amino acid residues present on the target Glucosyltransferase (GFT) of *Streptococcus mutans*. Based on the results of the computational analysis it was

concluded that the bio-active compound reveals significant binding against the target protein Glucosyltransferase (GFT) of *Streptococcus mutans*, thereby it was concluded that these compounds may exert promising to inhibit against GFT enzyme and hereby halt the catalytic transfer reactions which are essential for the survival of the organism *Streptococcus mutans*.

Periodontitis is caused by an inflammatory response to normal microbiota that is exacerbated by the presence of dysbiotic species. *P. gingivalis* is a "keystone pathogen" among these species, converting other benign biofilm members into pathobionts and causing aggressive damage to periodontal tissues.

Virulence factors of *P. gingivalis* include peptidases, which degrade proteins in infected tissues, nourishing bacteria and facilitating their spread and host colonisation. Peptidases also compromise host defences and outcompete bacterial competitors in periodontal pockets. The cysteine peptidases gingipain K are the most important (Kgp). Kgp is required for bacterial survival and the progression of periodontal disease, making it an ideal target for the development of new drugs [18].

Table 11. Summary of the Molecular Docking Studies of Compounds Against Prostaglandin H Synthases –PDB 1IGX

| Compounds | Binding Free Energy Kcal/mol | Inhibition Constant Ki μ M (*mM) (**nM) | Electrostatic Energy Kcal/mol | Intermolecular Energy Kcal/mol | Total Interaction Surface |
|------------|------------------------------|---|-------------------------------|--------------------------------|---------------------------|
| Vasicine | -5.34 kcal/mol | 121.37 μ M | -0.50 kcal/mol | -5.64 kcal/mol | 532.527 |
| Vasicinone | -5.66 kcal/mol | 70.64 μ M | -0.10 kcal/mol | -5.96 kcal/mol | 534.466 |
| Vasicoline | -5.75 kcal/mol | 60.71 μ M | -0.03 kcal/mol | -5.37 kcal/mol | 602.972 |
| Anisotine | -6.51 kcal/mol | 16.89 μ M | -0.00 kcal/mol | -7.08 kcal/mol | 630.438 |

Vasicoline and Anisotine revealed 100% binding efficacy by occupying the core amino acid residues (444 HIS, 477 CYS and 388 ASP) over the Gingipain K. Followed by compounds such as Vasicine and Vasicinone ranked second with the maximum of 2 interactions with the active site of the target enzyme Gingipain K. Vasicoline and Anisotine revealed significant binding affinity with all three active residual amino acids present on the active site of the target protein Gingipain K. Therefore, it was concluded that these compounds may exert promising inhibiting against Gingipain K.

The fimbriae are the main structures responsible for *P. gingivalis*'s virulent behaviour. They adhere to epithelial cells, fibroblasts, salivary components, and collagen, and thus play an important role in periodontal

tissue colonisation and invasion. The fim A gene encodes the main fimbria. Six genotypes of the FIM A gene have been identified based on the nucleotide sequence. Several studies have discovered links between periodontitis and a higher prevalence of fim A [19].

Nearly 24 amino acids are present in the sequence 339- 363 present in the active site of FIM A of *Porphyromonas gingivalis*. In our present investigation, it was observed that out of 4 compounds' the lead Anisotine reveals 3 potential interactions by occupying some of the active amino acid residues with the sequence (339- 363) present on the FIM A. Followed by this the compound Vasicine, Vasicinone and Vasicoline ranked second with the maximum of 1 interaction with the active site of the target protein FIM-A.

Anisotine revealed convincing binding affinity with the amino acids present on the active site of the target protein FIM-A and thereby it was concluded that the compound Anisotine may

TNF- has been linked to the immunology of periodontal disease. TNF- concentrations may be elevated in periodontal inflammation as a side effect of monocyte stimulation. This cytokine's elevation affects insulin sensitivity through both direct and indirect mechanisms, worsening diabetic status. A worsening of the diabetic condition may result in further periodontal breakdown. Thus, TNF- appears to play a key role in the vicious cycle that connects periodontal disease and diabetes [20].

exert some productive efficacy in hindering the bio-film formation by the organism *Porphyromonas gingivalis*.

Vasicinone, Vasicoline and Anisotine possessed a maximum of four interactions followed by Vasicine which had three viable interactions with the core active amino acid residues present on the target TNF-alpha. Therefore, all the bio-active compounds revealed significant binding affinity against the target enzyme TNF-alpha by interacting with active amino acid present on the active site thereby it was concluded that these compounds may exert promising anti-inflammatory activity.

Table 12. Amino Acid Residue Interaction of Lead Against Prostaglandin H Synthases –PDB 1IGX

| Compounds | Interaction | Amino acid Residues | | | | | | | | | | | | | | |
|----------------|-------------|---------------------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|
| | | | | | | | | | | | | | | | | |
| Vasicine | 4 | 205 PHE | 209 PHE | 344 VAL | 348 TYR | 349 VAL | 352 LEU | 381 PHE | 384 LEU | 385 TYR | 387 TRP | 530 SER | 534 LEU | | | |
| Vasicinone | 4 | 205 PHE | 344 VAL | 348 TYR | 349 VAL | 352 LEU | 381 PHE | 385 TYR | 530 SER | 534 LEU | | | | | | |
| Vasicoline | 3 | 344 VAL | 348 TYR | 349 VAL | 352 LEU | 381 PHE | 385 TYR | 518 PHE | 523 ILE | 527 ALA | 530 SER | | | | | |
| Anisotine | 3 | 116 VAL | 120 ARG | 348 TYR | 349 VAL | 352 LEU | 353 SER | 355 TYR | 381 PHE | 384 LEU | 385 TYR | 387 TRP | 523 ILE | 527 ALA | 530 SER | 531 LEU |
| Salicylic acid | 3 | 348 TYR | 349 VAL | 352 LEU | 381 PHE | 384 LEU | 385 TYR | 387 TRP | 518 PHE | 522 MET | 530 SER | | | | | |

PGE2 (prostaglandin E2) is a key inflammatory mediator in the pathophysiology of periodontitis. Three groups of enzymes act sequentially to catalyse PGE2 biosynthesis: phospholipase A2 (PLA2), cyclooxygenases (COX-1 and COX-2) and prostaglandin E (PGE) synthases, which act as catalysts in the final step of PGE2 synthesis. Therefore, inhibition of prostaglandin synthase H affects reducing periodontal inflammation [21].

Vasicoline, Anisotine, Vasicine and Vasicinone revealed a maximum of three to four interactions with the core active amino acid residues present on the target enzyme Prostaglandin H synthases. All the bio-active compounds revealed significant binding affinity against the target enzyme Prostaglandin H synthases by interacting with active amino acid present on the active site thereby it was concluded that these compounds may exert promising analgesic activity by inhibiting the activity of the enzyme Prostaglandin H synthases. It was concluded that the phytochemicals may act as potential therapeutic agents for the management of pain and inflammation.

From the above results it has been seen that Vasicine, Vasicinone, Vasicoline and Anisotine had a sufficient amount of Anti-

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Microbial activity towards cariogenic bacteria and periodontopathic bacteria. They also have analgesic and anti-inflammatory Activity. Therefore, they may find great use in the field of orthodontics.

Conclusion

The main point of the current research is that the compounds Vasicine, Vasicinone, Vasicoline, and Anisotine have significant binding affinity against various target enzymes related to cariogenic bacteria and periodontopathic bacteria, and they also possess analgesic and anti-inflammatory activity, making them potentially useful in the field of orthodontics. The said phytochemicals can further be explored for proper delivery to saliva, periodontal region and bone for effective use during and after orthodontic treatment.

Conflict of Interest

There is no conflict of interest.

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